ΓΕΝΙΚΑ ΘΕΜΑΤΑ

Βιολογική καταπολέμηση εχθρών

Νέες τάσεις και προκλήσεις σε ένα μεταβαλλόμενο περιβάλλον

Βιβλιογραφία

– Buitenhuis, R., Cock, M.J.W., Colmenarez, Y.C., De Clercq, P., Edgington, S., Gadaleta, P., Gwynn, R., Heimpel, G., Hill, M., Hinz, H.L., Hoddle, M.S., Jõkel, T., Klapwijk, J.N., Leung, K., Mc Kay, F., Messelink, G.J., Silvestri, L., Smith, D., Sosa, A., Wðckers, F.L., Cabrera Walsh, G., Wyckhuys, K.A.G. & Zaviezo, T. 2023. Sustainable use and conservation of microbial and invertebrate biological control agents and microbial biostimulants. Background Study Paper No. 71. Commission on Genetic Resources for Food and Agriculture. Rome, FAO. https://doi.org/10.4060/cb3571en

– Candolfi, M.P., Blumel, S., Foster, R., Bakker, F.M., Grimm, C., Hassan, S.A., Heimbach, U., MeadBriggs, M. A., Reber, B., Schmuck, R. & Vogt, H., eds. 2000. Guidelines to evaluate side-effects of plant protection products to non-target arthropods. Gent, Belgium, IOBC/WPRS

– Eilenberg, J., Hajek, A. & Lomer, C. 2001. Suggestions for unifying the terminology in biological control. BioControl, 46: 387–400.

– EPPO (European & Mediterranean Plant Protection Organization). 2014. Import and release of nonindigenous biological control agents. EPPO Bulletin, 44(3): 320–329. DOI: 10.1111/epp.12153

– EPPO Standards – PM 6 Safe use of biological control. 2023 https://www.eppo.int/RESOURCES/eppo_standards/pm6_biocontrol

– European Commission. 2020. A farm to fork strategy for a fair, healthy and environmentally-friendly food system. Brussels, European Commission. https://food.ec.europa.eu/horizontal-topics/farm-forkstrategy_en

– European Comission 2022. COMMISSION STAFF WORKING DOCUMENT Study on the Union's situation and options regarding the introduction, production, evaluation, marketing and use of invertebrate biocontrol agents within the territory of the Union. https://data.consilium.europa.eu/doc/document/ST-16317-2022-INIT/en/pdf

– FAO. 2017. Guidelines for the export, shipment, import and release of biological con https://www.fao.org/3/bt474e/ BT474E.pdf trol agents and other beneficial organisms.

– International Standard for Phytosanitary Measures 3. Rome. www.fao.org/3/j5365e/j5365e.pdf

– Garibaldi, L.A., Andersson, G.K., Requier, F., Fijen, T.P., Hipσlito, J., Kleijn, D., Pırez-Mındez, N. & Rollin, O. 2018. Complementarity and synergisms among ecosystem services supporting crop yield. Global Food Security, 17: 38–47. https://doi.org/10.1016/j.gfs.2018.03.006

– Gurr, G.M., Wratten, S.D. Landis, D.A. & You, M.S. 2017. Habitat management to suppress pest populations: progress and prospects. Annual Review of Entomology, 62: 91–107. DOI: 10.1146/annurevento-031616-035050

– Hajek, A.E., Gardescu, S. & Delalibera, I. 2021.Summary of classical biological control introductions of entomopathogens and nematodes for insect control. BioControl, 66: 167–180. https:// doi.org/10.1007/s10526-020-10046-7

– Happe, A.K., Alins, G., Blóthgena, N., Boreux, V., Bosch, J., Garcva, D., Hambðck, P.A. et al. 2019. Predatory arthropods in apple orchards across Europe: responses to agricultural management, adjacent habitat, landscape composition and country. Agriculture, Ecosystems & Environment, 273: 141–150. https://doi.org/10.1016/j.agee.2018.12.012

– Heimpel, G. & Cock, M.J.W. 2018. Shifting paradigms in the history of classical biological control. BioControl, 63: 27–37. https://doi.org/10.1007/ s10526-017-9841-9

– Heimpel, G.E. & Jervis, M.A. 2005. Does floral nectar improve biological control by parasitoids? In: F. Wðckers, P. vanRijn & J. Bruin, eds. Plant-provided food for carnivorous insects: a protective mutualism and its applications, pp. 367–304. Cambridge, UK,Cambridge University Press. https://doi.org/10.1017/CBO9780511542220

– Heimpel, G.E. & Mills, N.J. 2017. Biological control: ecology and applications. Cambridge, UK, Cambridge University Press. https://doi. org/10.1017/9781139029117

– Katsoyannos, P. (1996). Integrated Insect Pest Management for Citrus In Northern Mediterranean Countries. Benaki Phytopathological Institute, Athens, Greece

– Kenis, M., Hurley, B.P., Hajek, A.E & Cock, M.J.W. 2017.Classical biological control of insect pests of trees: facts and figures. Biological Invasions, 19: 3401–3417. https://doi.org/10.1007/s10530-017-1414-4

– Lampkin, N., Schwarz, G. & Bellon, S. 2020. Policies for agroecology in Europe, building on experiences in France, Germany and the United Kingdom. Landbauforschung – Journal of Sustainable and Organic Agricultural Systems, 70(2): 103–113. DOI: 10.3220/LBF1611684471000

– Schaffner, U., Knapp, M. & Seier, M. 2021. Biological control successes and failures: Eurasian region. In: P.G. Mason, ed. Biological control: global impacts, challenges and future directions of pest management, pp. 403–437. Clayton South, Australia, CSIRO Publishing. DOI: 10.1071/9781486309351

– Sheppard, A.W., Paynter, Q., Mason, P., Murphy,S., Stoett, P., Cowan, P., Brodeur, J. et al. 2019. The application of biological control for the management of established invasive alien species causing environmental impacts. IUCN SSC Invasive Species Specialist Group. Technical Series No. 91. Montreal, Canada, Secretariat of the Convention on Biological Diversity.

– Shields, M.W., Johnson, A.C., Pandey, S., Cullen, R., Gonzαlez-Chang, M., Wratten, S.D. & Gurr, G.M. 2019. History, current situation and challenges for conservation biological control. Biological control, 131: 25–35. https://doi.org/10.1016/j.biocontrol.2018.12.010

– Timper, P. 2014. Conserving and enhancing biological control of nematodes. Journal of Nematology, 46(2): 75–89. PMID: 24987159; PMCID: PMC4077175

– Torres, J.B. & Bueno, A.D.F. 2018. Conservation biological control using selective insecticides – a valuable tool for IPM. Biological Control, 126: 53–64. https://doi.org/10.1016/j.biocontrol.2018.07.012

– Van Driesche, R.G., Winston, R.L. & Duan, J.J. 2020. Classical insect biocontrol in North America, 1985 to 2018: a pest control strategy that is dying out? CAB Reviews, 15(37): 1–9. DOI: 10.1079/pavsnnr202015037

- Wade, M.R., Zalucki, M.P., Wratten, S.D. & Robinson, K.A. 2008. Conservation biological control of arthropods using artificial food sprays: current status and future challenges. Biological Control, 452(2): 185–199. https://doi.org/10.1016/j.biocontrol.2007.10.024
- Wratten, S.D., Gillespie, M., Decourtye, A., Mader, E. & Desneux, N. 2012. Pollinator habitat enhancement: Benefits to other ecosystem services. Agriculture, Ecosystems & Environment, 159: 112–122. https://doi. org/10.1016/j.agee.2012.06.020
- Ανώνυμος. Εθνικός Κατάλογος Μακρο-οργανισμών ΥΠΑΑΤ http://www.minagric.gr/index.php/el/for-farmer-2/crop-production/fytoprostasiamenu/skeyasmata-makroorganismoi Πρόσβαση 26/04/2022.
- Ν. ΠΕΤΡΟΠΟΥΛΟΣ, Η. ΓΕΩΡΓΟΠΟΥΛΟΥ, Π. ΜΥΛΩΝΑΣ, Δ.Π. ΠΑΠΑΧΡΗΣΤΟΣ, Ι. ΜΑΝΤΖΟΥΤΣΟΣ, Σ. ΜΠΟΥΚΟΥΒΑΛΑΣ, Γ. ΚΑΡΑΘΑΝΟΥ ΚΑΙ Δ. ΖΩΑΚΗ Εφαρμογή της κλασικής βιολογικής καταπολέμησης για το έντομο Metcalfa pruinosa (Hemiptera:Fulgoroidea) με την αξιοποίηση Γεωγραφικών Πληροφοριακών Συστημάτων. 16 Πανελλήνιο Εντομολογικό Συνέδριο, Ηράκλειο, 20-23 Οκτωβρίου 2015.
- Ανώνυμος (2021) Έκθεση Εργασιών Έτους 2021, Μπενάκειο Φυτοπαθολογικό Ινστιτούτο https://www.bpi.gr/files/ektheseis2021/%CE%95%CE%9A%CE%98%CE%95%CE%A3%CE%97%20%CE%95%CE%A1%CE%93%CE%91%CE%A3%CE%99%CE%A9%CE%9D%20

ΚΗΠΕΥΤΙΚΑ

Νέα δεδομένα για την αιτιολογία του ίκτερου της πιπεριάς στη χώρα μας

Εμπλοκή διαφορετικών ιών που μεταδίδονται με αλευρώδεις και μύκητες

Βιβλιογραφία

- Bos, L., and Huijberts, N. 1996. Lettuce ring necrosis, caused by a chytrid-borne agent distinct from lettuce big-vein 'virus'. European Journal of Plant Pathology, 102(9), 867-873. https://doi.org/10.1007/bf01877057
- Fletcher, J.T., Wallis, W.A., Davenport, F., 1987. Pepper yellow vein, a new disease of sweet peppers. Plant Pathology 36, 180-184. http://dx.doi.org/10.1111/j.1365-3059.1987.tb02219.x.
- Gavrili V., Leonidas Lotos, L., Katis, N. and Maliogka, V., 2021. First report of tomato chlorosis virus in pepper in Greece. Journal of Plant Pathology https://doi.org/10.1007/s42161-021-01022-w.
- Gambley, C., Persley, D. and Thomas, J.E., 2019. First record of Ranunculus white mottle virus from Australia. New Disease Reports (2019) 40, 13. http://dx.doi.org/10.5197/j.2044-0588.2019.040.013
- Lotos, L., Olmos, A., Orfanidou, C., Efthimiou, K., Avgelis, A., Katis, N.I. and Maliogka, V.I., 2017. Insights into the etiology of Polerovirus – induced Pepper Yellows Disease. Phytopathology 107(12):1567–1576. https:// doi. org/ 10. 1094/ PHYTO-07- 16- 0254-R
- Maachi, A., Torre, C., Sempere, R.N., Hernando, Y., Aranda, M.A. and Donaire, L., 2021. Use of High-Throughput Sequencing and Two RNA Input Methods to Identify Viruses Infecting Tomato Crops. Microorganisms 9, 1043. https://doi.org/10.3390/microorganisms9051043.
- Orfanidou, C.G., Dimitriou, C., Papayiannis, L.C., Maliogka, V.I. and Katis, N.I., 2014. Epidemiology and genetic diversity of criniviruses associated with tomato yellows disease in Greece. Virus Research 186: 120-129.
- Orfanidou, C.G., Pappi, P.G., Efthimiou, K.E., Katis, N.I. and Maliogka, V.I., 2016. Transmission of Tomato chlorosis virus (ToCV) by Bemisia tabaci biotype Q and evaluation of four weed species as viral sources. Plant Disease 100: 2043-2049.
- Rivarez, M. P. S., Kogej, Z., Jako , N., A. Pecman, A., Seljak, G., A. Vučurović, A., Ravnikar, M., Mehle, N. and Kutnjak, D., 2022. First Report of Ranunculus white mottle ophiovirus in Slovenia in pepper with yellow leaf curling symptom and in Tomato. Plant Disease 106:2003.
- Schoen, R., de Koning P.P.M., de Krom, C., Oplaat C., Westenberg M., Massart, S., Temple, C., De Jonghe, K., Blouin, A.G. and Botermans, M., 2023. First report of lettuce ring necrosis virus in chili pepper and tomato in Belgium and The Netherlands. Plant Disease https://doi.org/10.1094/PDIS-01-23-0036-PDN
- Vaira, A. M., Accotto, G. P., A. Costantini, A. and Milne, R. G. 2003. The partial sequence of RNA 1 of the ophiovirus Ranunculus white mottle virus indicates its relationship to rhabdoviruses and provides candidate primers for an ophiovirus-specific RT-PCR test. Archives of Virology 148: 1037–1050 DOI 10.1007/s00705-003-0016-x.
- Vicentin, E., Mituti, T., Nogueira, A.M., Fecury Moura, M. and Bello, V.H., Ribeiro-Junior, M.R., Wintermantel, W.M., Fiallo-Olive E., Navas-Castillo J., Krause-Sakate, R., Marques Rezende, J.A., 2022. Differential reaction of sweet pepper to infection with the crinivirus tomato chlorosis virus probably depends on the viral variant. Plant Pathology 71:1313–1322.

ΦΥΤΑ ΜΕΓΑΛΗΣ ΚΑΛΛΙΕΡΓΕΙΑΣ

Ανθεκτικότητα ζιζανίων σε ζιζανιοκτόνα στη χώρα και διεθνώς

Μέθοδοι και συστήματα διαχείρισης

ΒΙΒΛΙΟΓΡΑΦΊΑ (Διεθνής)

- Anthimidou, E., S. Ntoanidou, P. Madesis, and I. Eleftherohorinos. 2020. Mechanisms of Lolium rigidum multiple resistance to ALS- and ACCase-inhibiting herbicides and their impact on plant fitness. Pesticide Biochemistry and Physiology 164: 65-72.
- Barrθs, B., A. Micoud, M-F. Corio-Costet, D. Debieu, S. Fillinger, A-S Walker, C. Dılye, J. Grosman, and M. Siegwart. 2016. Trends and challenges in pesticide rsistance detection. Trends in Plant Science 21: 834-853.
- Beckie, H.J. and K.N. Harker. 2017. Our top 10 herbicide-resistant weed management practices. Pest Management Science 73: 1045-1052.
- Beckie, H.J. and F.J. Tardif. 2012. Herbicide cross-resistance in weeds. Crop Protection 35: 15-28.
- Delye, C. 2013. Unravelling the genetic bases of non-target-site-based resistance (NTSR) to herbicides: a major challenge for weed science in the forthcoming decade. Pest Management Science 69: 176-187.

- Gaines, T.A., S.O. Duke, S. Morran, C.A.G. Rigon, P.J. Tranel, A. Kópper, and F.E. Dayan. 2020. Mechanisms of evolved herbicide resistance. Journal of Biological Chemistry 295: 10307–10330; doi 10.1074/jbc.REV120.013572.
- Giannopolitis, C.N. and G. Vassiliou. 1989. Propanil tolerance in Echinochloa crus-galli (L.) Beauv. Tropical Pest Management 35: 6-7.
- Heap, I. 2023. International herbicide resistant weed database Available at web site https://www.weedscience.org/Home.aspx
- Kaloumenos, N.S., V.N. Adamouli, C.A. Dordas, and I.G. Eleftherohorinos. 2011. Corn poppy (Papaver rhoeas) cross-resistance to ALS-inhibiting herbicides. Pest Management Science 67: 574-585.
- Kaloumenos, N.S., N. Capote, A. Aguado and I.G. Eleftherohorinos. 2013a. Red rice (Oryza sativa) cross-resistance to imidazolinone herbicides used in resistant rice cultivars grown in northern Greece. Pesticide Biochemistry and Physiology 105: 177-183.
- Kaloumenos, N.S., S.L. Chatzilazaridou, P.V. Mylona, A.N. Polidoros, and I.G. Eleftherohorinos. 2013b. Target-site mutation associated with cross-resistance to ALS-inhibiting herbicides in late watergrass (Echinochloa oryzicola Vasing). Pest Management Science 69: 865-873.
- Kaloumenos, N.S., C.A. Dordas, G.C. Diamantidis, and I.G. Eleftherohorinos. 2009. Multiple Pro197 substitutions in the acetolactate synthase of corn poppy (Papaver rhoeas) confer resistance to tribenuron. Weed Science 57: 362-368.
- Kaloumenos, N.S. and I.G Eleftherohorinos. 2008. Corn poppy (Papaver rhoeas) resistance to ALS-inhibiting herbicides and its impact on growth rate. Weed Science 56: 789-796.
- Kaloumenos, N.S. and I.G. Eleftherohorinos. 2009. Identification of a johnsongrass (Sorghum halepense) biotype resistant to ACCase-inhibiting herbicides in northern Greece. Weed Technology 23: 470-476.
- Kaloumenos, N.S., V.C. Tsioni, E.G. Daliani, S.E. Papavassileiou, A.G. Vassileiou, P.N. Laoutidou, I.G. Eleftherohorinos. 2012. Multiple Pro-197 substitutions in the acetolactate synthase of rigid ryegrass (Lolium rigidum) and their impact on chlorsulfuron activity and plant growth. Crop Protection 38: 35-43.
- Kanatas, P., A. Tataridas, V, Dellaportas, and I. Travlos. 2021. First report of Amaranthus palmeri S. Wats. in cotton, maize and sorghum in Greece and problems with its management. Agronomy 11, 1721; doi.org/10.3390/agronomy11091721
- Kati V., L. Scarabel, D. Thiery-Lanfranchi, V. Kioseoglou, S. Liberopoulou and C. Delye. 2019. Multiple resistance of Papaver rhoeas L. to 2,4-D and acetolactate synthase inhibitors in four European countries. Weed Research 59: 367–376.
- Mylonas, P.N., C.N. Giannopolitis, P.G. Efthimiadis, G. Menexes, P.B. Madesis, I.G. Eleftherohorinos. 2014. Glyphosate resistance of molecularly identified Conyza albida and Conyza bonariensis populations. Crop Protection 65: 207-215.
- Mylonas, P.N., C.N. Giannopolitis, P.G. Efthimiadis, G.C. Menexes, G.C., and I.G. Eleftherohorinos. 2019. Dose-response and growth rate variation among glyphosate resistant and susceptible Conyza albida and Conyza bonariensis populations. Journal of Plant Protection Research 59: 32-40.
- Ntoanidou, S., N. Kaloumenos, G. Diamantidis, P. Madesis, I. Eleftherohorinos. 2016. Molecular basis of Cyperus difformis cross-resistance to ALS-inhibiting herbicides. Pesticide Biochemistry and Physiology 127: 38-45.
- Ntoanidou, S., P. Madesis, G. Diamantidis, I. Eleftherohorinos. 2017. Trp574 substitution in the acetolactate synthase of Sinapis arvensis confers cross-resistance to tribenuron and imazamox. Pesticide Biochemistry and Physiology 142: 9–14.
- Ntoanidou, S., P. Madesis, I. Eleftherohorinos. 2019. Resistance of Rapistrum rugosum to tribenuron and imazamox due to Trp574 or Pro197 substitution in the acetolactate synthase. Pesticide Biochemistry and Physiology 154: 1-6.
- Ntoanidou, S., P. Madesis, G. Menexes, and I. Eleftherohorinos. 2020. Growth rate and genetic structure of Sinapis arvensis susceptible and herbicide resistant populations originating from Greece. Euphytica 216 (12): 185.
- Papapanagiotou, A., I. Bosmali, P. Madesis, G. Menexes and I. Eleftherohorinos. 2022a. Multiple-resistance of silky windgrass to acetolactate synthase (ALS)- and acetyl-CoA synthase (ACCase)-inhibiting herbicides. Weed Technology 36: 334-343; doi: 10.1017/wet.2022.24.
- Papapanagiotou, A., I. Bosmali, P. Madesis, G. Menexes and I. Eleftherohorinos. 2022. Multiple-resistance of silky windgrass to acetolactate synthase (ALS)- and acetyl-CoA synthase (ACCase)-inhibiting herbicides. Weed Technology 36: 334-343. Doi: 10.1017/wet.2022.24.
- Papapanagiotou, A.P., C.A. Damalas, I. Bosmali, P. Madesis, G.C. Menexes, I.G. Eleftherohorinos. 2019. «False cleavers (Galium spurium L.) and common cleavers (G. aparine L.) resistance to ALS-inhibiting herbicides in northern Greece». Planta Daninha v37:e019207288.
- Papapanagiotou, A.P., C.A. Damalas, G.C. Menexes, I.G. Eleftherohorinos. 2020. Resistance levels and chemical control options of sterile oat (Avena sterilis L.) in Northern Greece. International Journal of Pest Management 66: 106-115.
- Papapanagiotou, A.P., N.S. Kaloumenos, and I.G. Eleftherohorinos. 2012. Sterile oat (Avena sterilis L.) cross-resistance profile to ACCase-inhibiting herbicides in Greece. Crop Protection 35: 118-126.
- Papapanagiotou, A., D. Loukovitis, S. Ntoanidou and I. Eleftherohorinos. 2022b. Target-site cross-resistance to ALS inhibitors in johnsongrass [Sorghum halepense (L.) Pers.] originating from Greek corn fields. Weed Technology 36: 276–282; doi: 10.1017/wet.2022.8
- Papapanagiotou, A.P., D. Loukovitis, E. Anthimidou, and I.G. Eleftherohorinos. 2023. Impact of ALS-herbicide resistant perennial ryegrass (Lolium perenne L.) population on growth rate and competitive ability against wheat. (Submitted for publication).
- Papapanagiotou, A., D. Loukovitis, C. Damalas and I. Eleftherohorinos. 2022c. Identification of an ACCase-resistant johnsongrass (Sorghum halepense L.) population from a cotton field in northern Greece. Weed Biology and Management 22: 88–93; doi.org/10.1111/wbm.12256.
- Papapanagiotou, A.P., M.I. Paresidou, N.S. Kaloumenos, and I.G. Eleftherohorinos. 2015. ACCase mutations in Avena sterilis

populations and their impact on plant fitness. Pesticide Biochemistry and Physiology 123: 40-48.
- Papapanagiotou, A., Th. Spanos N.E. Zarrougui, I.C. Livieratos, and I.G. Eleftherohorinos. 2023. Pro197 and Trp574 substitutions in the acetolactate synthase of corn marigold and their impact on competitive ability against barley. Weed Technology, doi: 10.1017/wet.2023.17.
- Papapanagiotou, A., I. Vasilakoglou, K. Dhima, and I. Eleftherohorinos. 2023. Growth rate and competitive ability of susceptible and multiple-resistant late watergrass (Echinochloa phyllopogon) biotypes to rice. Phytoparasitica (in press).
- Scarabel L, S. Panozzo, D. Loddo, SK Mathiassen, M. Kristensen, P. Kudsk, T. Gitsopoulos, I. Travlos, E. Tani, D. Chachalis and M. Sattin. 2020. Diversified resistance mechanisms in multi-resistant Lolium spp. in three European countries. Frontiers Plant Science 11:608845; doi: 10.3389/fpls.2020.608845.
- Travlos, I., I. Tabaxi, D. Papadimitriou, D. Bilalis and D. Chachalis. 2016. Lolium rigidum Gaud. biotypes from Greece with resistance to glyphosate and other herbicides. Bulletin of University of Agricultural Sciences and Veterinary Medicine Cluj-Napoca. Horticulture 73(1); doi: 10.15835/buasvmcn-hort:11772.
- Travlos, I.S., N. Cheimona , R. De Prado 2 , A.J. Jhala , D. Chachalis , and E. Tani. 2018. First Case of Glufosinate-Resistant Rigid Ryegrass (Lolium rigidum Gaud.) in Greece . Agronomy 8(4) 35; doi.org/10.3390/agronomy8040035.
- Travlos, I.S. and D. Chachalis. 2010. Glyphosate-resistant hairy fleabane (Conyza bonariensis) is reported in Greece. Weed Technology 24: 569-573.
- Travlos, I., P. Kanatas, A. Tsekoura, I. Gazoulis, P. Papastylianou, I. Kakabouki, and N. Antonopoulos. 2020. Efficacy of different herbicides on Echinochloa colona (L.) Link control and the first case of Its glyphosate resistance in Greece. Agronomy, 10, 1056; doi: 10.3390/agronomy10071056.
- Vasilakoglou, I.B., I.G. Eleftherohorinos and K.V. Dhima. 2000. Propanil-resistant barnyardgrass (Echinochloa crus-galli) biotypes found in Greece. Weed Technology 14: 524-529.
- Vasilakoglou, I., K. Dhima and T. Gitsopoulos. 2018. Management of penoxsulam- and bispyribac-resistant late watergrass (Echinochloa phyllopogon) biotypes and rice sedge (Cyperus difformis) in rice. Chilean Journal of Agricultural Research 78: 2; doi.org/10.4067/S0718-58392018000200276.

ΒΙΒΛΙΟΓΡΑΦΙΑ (εθνική)

- Ελευθεροχωρινός, Η. Γ. 2020. Ζιζανιολογία: Βιολογία και Διαχείριση Ζιζανίων-Ζιζανιοκτόνα, Φυτά και Περιβάλλον. Εκδόσεις Αγροτύπος Α.Ε. Αθήνα. 5η Έκδοση, 497 σελ.
- Ελευθεροχωρινός, Η.Γ. και Δ.Μ. Παπαμιχαήλ. 2002. Ανάπτυξη ανθεκτικού βιοτύπου αγριοτοματιάς (Solanum nigrum L.) στο ζιζανιοκτόνο prometryn. 12ο Επιστημονικό Συνέδριο Ελληνικής Ζιζανιολογικής Εταιρίας. Αθήνα 2-3 Δεκεμβρίου 2002. Περιλήψεις Ανακοινώσεων σελ. 68.
- Μυλωνάς, Φ., Β. Κατή, Η. Ελευθεροχωρινός. 2017. Αξιολόγηση πληθυσμών Avena sterilis και Galium spp. για ανθεκτικότητα σε ζιζανιοκτόνα-αναστολείς της δράσης των ενζύμων ALS/ACCase και ALS. 19ο Επιστημονικό Συνέδριο, Νέα Ορεστιάδα 29-31 Μαρτίου 2017.
- Παπαπαναγιώτου, Α., Γ. Μενεξές, Η. Ελευθεροχωρινός. 2017a. Πληθυσμοί μουχρίτσας πολλαπλώς ανθεκτικοί σε ζιζανιοκτόνα-αναστολείς των ενζύμων ALS/ACCase. 19ο Επιστημονικό Συνέδριο, Νέα Ορεστιάδα 29-31 Μαρτίου 2017.
- Παπαπαναγιώτου, Α., Γ. Μενεξές, Η. Ελευθεροχωρινός. 2017b. Πληθυσμοί μικρόκαρπης κολλητσίδας και μπιφόρας ανθεκτικοί σε ζιζανιοκτόνα-αναστολείς του ενζύμου ALS. 19ο Επιστημονικό Συνέδριο, Νέα Ορεστιάδα 29-31 Μαρτίου 2017.
- Παπαπαναγιώτου, Α., Γ. Μενεξές, Η. Ελευθεροχωρινός. 2017c. Πληθυσμοί αγριοβρώμης με διασταυρωτή ανθεκτικότητα σε ζιζανιοκτόνα-αναστολείς του ενζύμου ALS και μελέτη της προσαρμοστικότητάς τους. 19ο Επιστημονικό Συνέδριο, Νέα Ορεστιάδα 29-31 Μαρτίου 2017.
- Παπαπαναγιώτου, Α., Γ. Μενεξές, Η. Ελευθεροχωρινός. 2017d. Πλυθυσμοί μίλιου και ανεμόχορτου ανθεκτικοί σε ζιζανιοκτόνα-αναστολείς των ενζύμων ACCase και ALS. 19ο Επιστημονικό Συνέδριο, Νέα Ορεστιάδα 29-31 Μαρτίου 2017.
- Παπαπαναγιώτου, Α., Ι. Βασιλάκογλου, Κ. Δήμας, Η. Ελευθεροχωρινός. 2017e. Διερεύνηση της ανάπτυξης διασταυρούμενης ανθεκτικότητας του Sinapis arvensis και της ευαισθησίας του Camellina microcarpa σε ζιζανιοκτόνα-αναστολείς της δράσης του ενζύμου οξικογαλακτική συνθάση. 19ο Επιστημονικό Συνέδριο, Νέα Ορεστιάδα 29-31 Μαρτίου 2017.